

Monoamine Oxidase-Catalyzed Oxidative Decarboxylation of *cis*- and *trans*-5-Aminomethyl-3-(4-methoxyphenyl)dihydrofuran-2(3*H*)-one. Evidence for the Intermediacy of an α -Radical

Richard B. Silverman,* Joseph J. P. Zhou, Charles Z. Ding, and Xingliang Lu

Department of Chemistry and Department of Biochemistry, Molecular Biology, and Cell Biology Northwestern University, Evanston, Illinois 60208-3113

Received June 23, 1995

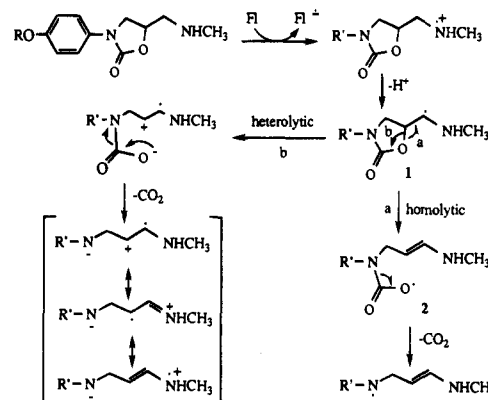
Monoamine oxidase (EC 1.4.3.1, MAO) catalyzes the oxidation of a variety of primary, secondary, and tertiary amines to give the corresponding imines.¹ Most of the mechanistic studies support a radical mechanism.^{2,3} Previously, we found that a class of oxazolidinone inactivators of MAO underwent MAO-catalyzed excision of CO₂ from the oxazolidinone ring by a route that did not lead to enzyme inactivation.⁴ Two mechanisms were proposed to account for this loss of CO₂, both proceeding from an α -radical (**1**, Scheme 1). Homolytic C–O bond cleavage (pathway a) leads to a carboxyl radical (**2**), which is known to undergo rapid decarboxylation.⁵ However, because of anion stabilization by the oxazolidinone nitrogen, decarboxylation also could have arisen from a heterolytic elimination route (pathway b).

The corresponding lactone also was shown to be a time-dependent inactivator of MAO,⁶ whose inactivation mechanism was hypothesized to be the result of formation of a covalent adduct at the α -position to the enzyme; the electron-withdrawing effect of the lactone ring was proposed to be responsible for adduct stabilization.⁷

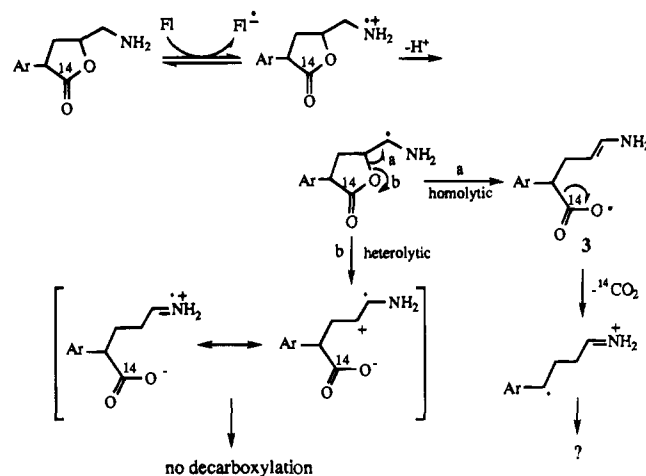
It occurred to us that excision of CO₂ from the lactone also should be possible, but only if an α -radical is generated and only if homolytic C–O bond cleavage occurs (Scheme 2, pathway a). As in the case of the oxazolidinone, this would generate a carboxyl radical (**3**), which is known to undergo rapid decarboxylation to the carbon radical.⁵ Heterolytic C–O bond cleavage from the α -radical (Scheme 2, pathway b), however, would produce a reactive species without loss of CO₂ because of the stability of alkyl carboxylates. α -Carbanion formation, by any mechanism, would lead to heterolytic C–O bond cleavage to give the carboxylate, which, also, would not undergo decarboxylation (Scheme 3). Loss of CO₂, therefore, is a signal for both α -radical formation and homolytic C–O bond cleavage.

To determine if CO₂ excision occurs during MAO-catalyzed oxidation of the lactone, [2-¹⁴C]-*cis*- (**4**) and [2-¹⁴C]-*trans*-5-aminomethyl-3-(4-methoxyphenyl)dihydrofuran-2(3*H*)-one (**5**) were synthesized by the route shown in Scheme 4.⁸ Incubation

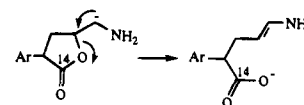
Scheme 1. Homolytic versus Heterolytic Mechanisms for Excision of CO₂ from Oxazolidinones



Scheme 2. Homolytic versus Heterolytic Mechanisms for MAO-Catalyzed Oxidation of 5-Aminomethyl-3-(4-methoxyphenyl)dihydrofuran-2(3*H*)-ones



Scheme 3. Fate of a Carbanion Intermediate



of homogeneous beef liver MAO with **4** and **5** resulted in the production of 0.5 and 6 equiv of ¹⁴CO₂, respectively,⁹ per inactivation event. Since homolytic C–O bond cleavage depends upon appropriate overlap between the orbital of the

(1) (a) Krueger, M. J.; Efang, S. M. M.; Michelson, R. H.; Singer, T. P. *Biochemistry* **1992**, *31*, 5611–5615. (b) Kalgutkar, A. S.; Castagnoli, N., Jr. *J. Med. Chem.* **1992**, *35*, 4165–4174. (c) Zhao, Z.; Dalvie, D.; Naiman, N.; Castagnoli, K.; Castagnoli, N., Jr. *J. Med. Chem.* **1992**, *35*, 4473–4478. (d) Yu, P. H.; Davis, B. A.; Boulton, A. A. *J. Med. Chem.* **1992**, *35*, 3705–3713. (e) Kalgutkar, A. S.; Castagnoli, K.; Hall, A.; Castagnoli, N., Jr. *J. Med. Chem.* **1994**, *37*, 944–949. (f) Ding, C. Z.; Nishimura, K.; Silverman, R. B. *J. Med. Chem.* **1993**, *36*, 1711–1715.

(2) Silverman, R. B. *Acc. Chem. Res.* **1995**, *28*, 335–342.

(3) Jonsson, T.; Edmondson, D. E.; Klinman, J. P. *Biochemistry* **1994**, *33*, 14871–14878.

(4) Gates, K. S.; Silverman, R. B. *J. Am. Chem. Soc.* **1990**, *112*, 9364–9372.

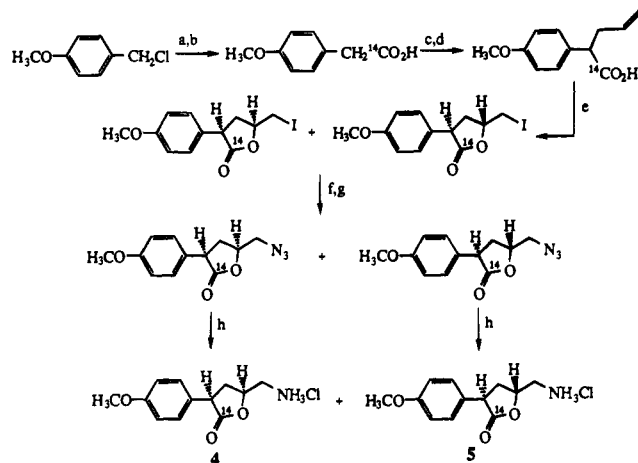
(5) (a) Schäfer, H. J. *Angew. Chem., Int. Ed. Engl.* **1981**, *20*, 911–934. (b) Fry, A. J. *Synthetic Organic Electrochemistry*, 2nd ed.; Wiley: New York, 1989. (c) Barton, D. J. R. *Aldrichim. Acta* **1990**, *23*, 3–10.

(6) Ding, Z.; Silverman, R. B. *J. Med. Chem.* **1992**, *35*, 885–889.

(7) Silverman, R. B.; Ding, C. Z. *J. Am. Chem. Soc.* **1993**, *115*, 4571–4576.

(8) Starting from 1 mCi of K¹⁴CN on a 1.0 mmol scale, 40 mg of **4** and 21 mg of **5** were obtained radiopure with specific activities of 1.0 mCi/mmol.

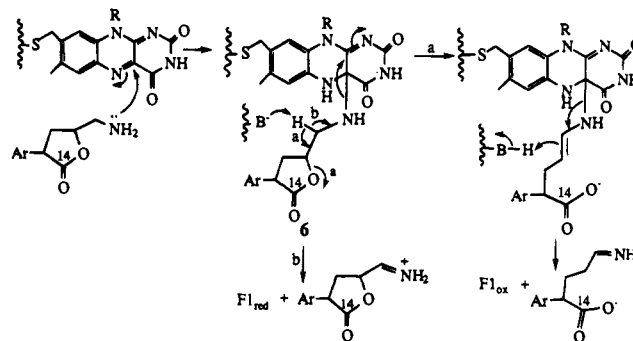
(9) In the case of **4**, MAO B (50 μ L, 199 μ M) in 100 mM sodium phosphate buffer, pH 7.4, was added to a 20 mL scintillation vial, which was stoppered with a serum cap with a Kontes center well (Cat. No. 882320-0000) in it. After 150 μ L of KOH (8.75%) was added to the center well, a solution of **4** in 100 mM sodium phosphate buffer, pH 7.4 (1.0 mL; **4** was dissolved in 100 μ L of DMSO and diluted in the buffer to give a solution containing 10% DMSO), was added to the bottom of the vial by syringe to give a final concentration of 4 mM. After the vial was shaken for 49.5 h at ambient temperature, 70% of the MAO was inactivated (relative to a control without **4**). Sulfuric acid (100 μ L, 3 M) was injected into the enzyme mixture. After being shaken for 30 min, the center well was removed and counted in 10 mL of scintillation fluid (toluene/dioxane/methanol/PPO-POPOP, 162.5/25.2/81.3/11.7). The total radioactivity was 5672 cpm (6752 dpm). Radioactivity trapped in a nonenzymatic control reaction, run in parallel (70 cpm; 87 dpm), was subtracted from the experiment. The number of equivalents of CO₂ produced was determined by inactivating MAO B with [³H]pargyline and setting that amount of radioactivity to 1 equiv. There was no difference in the result when the addition of sulfuric acid to quench the reaction and release the carbon dioxide was omitted from the procedure. A similar experiment was carried out with **5**, except that 109 μ M MAO B was used, and the inactivator was incubated with the enzyme for 60 h; total radioactivity trapped was 41 145 cpm (46 200 dpm), and the control was 18 cpm (20 dpm). The extended incubation times were needed because of the poor substrate/inactivator activities of **4** and **5**.

Scheme 4. Synthetic Routes to [2-¹⁴C]-*cis*- (**4**) and [2-¹⁴C]-*trans*-5-Aminomethyl-3-(4-methoxyphenyl)-dihydrofuran-2(3*H*)-one (**5**)^a

^a (a) K¹⁴CN/MeCN/18-C-6. (b) NaOH/EtOH. (c) *n*-BuLi. (d) Allyl bromide. (e) I₂/MeCN. (f) NaN₃/DMF. (g) Chromatography. (h) H₂/10% Pd-C/HCl.

C—O bond and the orbital containing the α -radical, and since **4** and **5** are diastereomers, a different amount of ¹⁴CO₂ generated from each is to be expected. Normal oxidation of these compounds to the corresponding aldehydes was determined by measuring the formation of radioactively-labeled nonamines produced, to get an indication of the relative incidence of decarboxylation versus amine oxidation (normal turnover). Compound **4** produced 146 equiv (total of 5.85×10^5 dpm) of nonamines, and **5** produced 281 equiv (total of 5.96×10^5 dpm).¹⁰ These results indicate that decarboxylation does not occur with a high percentage of the turnovers, but this is a function of the lifetime of the radical, the orientation of the C—O bond relative to the radical-containing orbital, and the rate of C—O bond cleavage relative to second-electron oxidation. Every turnover may proceed via the α -radical, but because of

(10) Different amounts of enzyme were used for these experiments: **4**, 1.73 nmol; **5**, 0.92 nmol.

Scheme 5. Fate of a Nucleophilic Mechanism for MAO-Catalyzed Oxidation of 5-Aminomethyl-3-(4-methoxyphenyl)dihydrofuran-2(3*H*)-ones

the above-mentioned factors, only a small percentage of the molecules may lead to decarboxylation.

These results also are further evidence against a nucleophilic mechanism for MAO.¹¹ Nucleophilic addition of **4** or **5** to the flavin would generate a covalent adduct (**6**, Scheme 5) that would not lead to loss of ¹⁴CO₂; deprotonation would lead either to elimination of the carboxylate and possible formation of a ¹⁴C metabolite (pathway a) or to elimination of the reduced flavin and a ¹⁴C metabolite (pathway b). No reasonable mechanism can account for generation of ¹⁴CO₂ from this covalent adduct.

In summary, MAO-catalyzed production of ¹⁴CO₂ from both [2-¹⁴C]-**4** and [2-¹⁴C]-**5** is strong evidence for the intermediacy of an α -radical and for subsequent (or concomitant) homolytic C—O bond cleavage. A nucleophilic mechanism is not consistent with these results.

Acknowledgment. The authors are grateful to the National Institutes of Health (GM32634) for financial support of this research.

JA952050N

(11) (a) Kim, J.-M.; Bogdan, M. A.; Mariano, P. S. *J. Am. Chem. Soc.* **1993**, *115*, 10591–10595. (b) Kim, J.-M.; Hoegy, S. E.; Mariano, P. S. *J. Am. Chem. Soc.* **1995**, *117*, 100–105.